

Available online at www.sciencedirect.com



**IL FARMACO** 

IL FARMACO 59 (2004) 33–39

www.elsevier.com/locate/farmac

# Synthesis of new 2,3-diaryl-1,3-thiazolidin-4-ones as anti-HIV agents

Angela Rao<sup>a</sup>, Jan Balzarini <sup>b</sup>, Anna Carbone<sup>a</sup>, Alba Chimirri<sup>a</sup>, Erik De Clercq<sup>b</sup>, Anna Maria Monforte<sup>a</sup>, Pietro Monforte<sup>a,\*</sup>, Christophe Pannecouque <sup>b</sup>, Maria Zappalà<sup>a</sup>

*<sup>a</sup> Dipartimento Farmaco-Chimico, Università di Messina, viale Annunziata, Messina 98168, Italy <sup>b</sup> Rega Institute for Medical Research, Katholieke Universiteit Leuven, 10 Minderbroedersstraat, Leuven B-3000, Belgium*

Received 26 July 2003; accepted 5 September 2003

# **Abstract**

Several 2,3-diaryl-1,3-thiazolidin-4-ones were synthesized and evaluated as anti-HIV agents. The results of the in vitro tests showed that some of them were highly effective inhibitors of HIV-1 replication at 30–50 nM concentrations with minimal cytotoxicity, thereby acting as non-nucleoside HIV-1 reverse transcriptase inhibitors (NNRTIs).

© 2003 Éditions scientifiques et médicales Elsevier SAS. All rights reserved.

*Keywords:* 2,3-diaryl-1,3-thiazolidin-4-ones; Anti-HIV activity; NNRTIs

# **1. Introduction**

The therapeutic agents currently approved for the treatment of HIV-1 infections include three non-nucleoside reverse transcriptase inhibitors (NNRTIs): nevirapine, delavirdine and efavirenz [\[1\].](#page-5-0) Unlike the nucleosides/nucleotides that act at the catalytic site of HIV reverse transcriptase (RT) by terminating DNA synthesis, NNRTIs bind in a region of the enzyme, which is approximately 10  $\AA$  away from the catalytic site. Their binding appears to result in a distortion of the catalytic site, which affects the ability of the enzyme to carry out its catalytic functions [\[2\].](#page-5-0)

Since they are not analogues of natural compounds and do not utilize the biochemical machinery of the host cells, NNR-TIs usually manifest side effects milder than those resulting from treatment with nucleosides. Although the therapeutic potential of NNRTIs has been compromised by the rapid development of resistance, they have proven to be useful in combination therapy with nucleoside RT and protease inhibitors [\[3\].](#page-5-0)

A multiple-drug treatment approach avoids or delays emergence of resistant viral strains and has indeed contributed to the declining morbidity and mortality among HIVinfected patients [\[4\].](#page-5-0) However, there are certain factors that

\* Corresponding author.

*E-mail address:* monforte@pharma.unime.it (P. Monforte).

restrict the selection of the agents for combination therapy, including drug compatibilities, adverse effects and cross resistance. Therefore, the synthesis of new effective NNRTIs remains a worthwhile goal.

In the course of our studies aimed at the discovery of new NNRTIs [\[5–7\],](#page-5-0) we have found that members of a series of 2,3-diaryl-1,3-thiazolidin-4-ones [\[8–12\]](#page-6-0) were highly effective in inhibiting the cytopathic effect of HIV-1 in human T-lymphocyte cells. The RT activity inhibition assays demonstrated that these compounds represent a new class of NNRTIs [\[8\].](#page-6-0) Preliminary molecular modelling studies suggested that the binding site of these compounds, similarly to other NNRTIs, is the non-nucleoside-binding pocket [\[10\].](#page-6-0)

We have demonstrated that a high activity level was associated with the presence of a 2,6-dihalosubstituted phenyl ring at C-2. Moreover, we found that an increase in antiviral potency was dependent on the presence of a (hetero)aromatic nucleus at N-3 bearing a small lipophilic substituent at the *meta* position.

Following these results and in order to better determine the structural characteristics able to improve the anti-HIV-1 activity of this class of compounds, we extended our studies to the synthesis of a new series of 1,3-thiazolidin-4-ones in which the optimal (hetero)aromatic ring at N-3 was maintained, whereas the 2,6-dihalophenyl ring at C-2 was modified by varying the substitution pattern in terms of nature and number of the substituents.

<sup>© 2003</sup> Éditions scientifiques et médicales Elsevier SAS. All rights reserved. doi:10.1016/j.farmac.2003.09.001

Compounds synthesized were evaluated for prevention of the cytopathic effect of HIV in cell-based assays. In addition, to confirm their mechanism of action, the inhibitory effects on HIV-1 RT were examined.

## **2. Results and discussion**

The synthesis of the new 2,3-diaryl-1,3-thiazolidin-4 ones was carried out by reacting a properly-substituted benzaldehyde with an equimolar amount of a suitable (hetero)aromatic amine in the presence of an excess of mercaptoacetic acid in refluxing toluene (Scheme 1). The products obtained were isolated by conventional workup in satisfactory yields. Both analytical and <sup>1</sup>H NMR spectral data of all the synthesized compounds are in full agreement with the proposed structures.

All compounds obtained were tested for anti-HIV activity by determining their ability to inhibit the replication of HIV-1  $(III_B)$  or HIV-2 (ROD) in human T-lymphocyte (MT-4) cells and the results are reported in [Table 1,](#page-2-0) in which the data of some 2-(2,6-difluorophenyl)-derivatives (i.e. **1, 7, 13, 19, 25** and **31**) have been included for comparison purposes. The anti-HIV activity of several compounds was also measured in HIV-1  $(III_B)$  infected CEM cells [\(Table 2\)](#page-3-0). Compound-induced cytotoxicity was also measured in MT-4 and CEM cells parallel with the antiviral activity. A select group of compounds was moreover evaluated for the inhibitory effects on HIV-1 RT enzymatic activity and the results are also reported in [Table 2.](#page-3-0)

Several of the new compounds prevented the cytopathic effect of HIV-1  $III<sub>B</sub>$  at nanomolar concentrations and were minimally toxic to MT-4 or CEM cells resulting in remarkably high selectivity indices. It is worth noting that compound **24**, one of the most promising of the series, possessed a selectivity index >7000.

As observed for other classes of NNRTIs, none of the tested compounds inhibited the replication of HIV-2 (ROD) in MT-4 cells at subtoxic concentrations (data not shown).

The in vitro  $IC_{50}$  values for HIV-1 RT with poly(rC).oligo(dG) as the template/primer were significantly higher than the corresponding  $EC_{50}$  values for inhibition of the cytopathic effect of HIV-1 in MT-4 cell culture. This discrepancy is not unusual for NNRTIs as it may reflect the differences between the in vitro HIV-1 RT assay, in which a synthetic template/primer is used, and the cellular systems [\[13\].](#page-6-0)

From the structure–activity relationship point of view, the antiviral activity varies considerably with the nature of both (hetero)aromatic nucleus at N-3 position and the substituents on the phenyl ring at C-2. In fact, pyridin-2-yl derivatives are more active than the corresponding pyrimidin-2-yl and phenyl ones. It is also worth noting that in the pyridin-2-yl series a bromine atom or a methyl group at 6-position is an important feature for potent anti-HIV agents and that the shift of the methyl group from 6- to 4-position negatively influences the activity. Considering the substitution pattern on the C-2 phenyl ring, the results obtained confirm that the presence of two halogen atoms at 2- and 6-positions is of paramount importance to increase the activity. Indeed, as previously reported [\[12\],](#page-6-0) these structural features restrict the rotation of the phenyl ring and allow the molecules to assume the characteristic butterfly-like conformation present in many other known NNRTIs [\[14\].](#page-6-0) The replacement of one fluorine atom with a methoxy group maintains the activity (**21** vs. **19, 33** vs. **31**), whereas the presence of two methoxy groups induces a decrease in activity with the sole exception of **24** which is one of the most promising derivatives of the series.

Furthermore, the substitution of both fluorine atoms with methyl groups is detrimental for anti-HIV activity leading to loss of potency of one order of magnitude.

Of the 2,3,5-trisubstituted congeners, the best results have been recorded among the 2-chloro,3-methyl,6-fluoro derivatives, the most active of which is compound **17** that possesses an  $EC_{50} = 0.050 \mu M$ .

In conclusion, a new series of 2,3-diaryl-1,3-thiazolidin-4-ones was synthesized and characterized and some of them proved to be potent anti-HIV agents. Once again, the results reported in this study confirm that the anti-HIV activity in this class of NNRTIs is strongly dependent on the nature of the substituents at C-2 and N-3 of the thiazolidinone ring.

# **3. Experimental**

#### *3.1. Chemistry*

Melting points were determined on a Kofler hot-stage apparatus and are uncorrected. Elemental analyses (C, H and N) were carried out on a Carlo Erba Model 1106 Elemental Analyzer and the results are within ±0.4% of the



Scheme 1. Synthesis of 2,3-diaryl-1,3-thiazolidin-4-ones.

<span id="page-2-0"></span>





<sup>a</sup> 50% Effective concentration or concentration required to reduce HIV-1-(III<sub>B</sub>)-induced cytopathic effect by 50% in MT-4 cells. b Cytotoxic concentration or concentration required to reduce MT-4 cell viability by 50%.

<sup>c</sup> Selectivity index: ratio of CC<sub>50</sub> to EC<sub>50</sub>.<br>d Data from Ref. [\[11\].](#page-6-0)

<sup>e</sup> Data from Ref. [\[10\].](#page-6-0)

<sup>f</sup> Data from Ref. [\[12\].](#page-6-0) NA, not active.

theoretical values. Merck silica gel 60  $F_{254}$  plates were used for analytical TLC; column chromatography was performed on Merck silica gel 60 (230–400 mesh). <sup>1</sup>H NMR spectra were recorded in CDCl<sub>3</sub> on a Varian Gemini-300 spectrometer. Chemical shifts were expressed in  $\delta$  (ppm) relative to TMS as internal standard and coupling constants (*J*) in Hz.

# *3.1.1. General procedure for the synthesis of 2,3-diaryl-1,3-thiazolidin-4-ones*

To a stirred solution of the aromatic amine (8 mmol) in dry toluene (50 ml), 2-mercaptoacetic acid (16 mmol) and the appropriate aldehyde (8 mmol) were added. The reaction mixture was refluxed for 48 h and then neutralized by a <span id="page-3-0"></span>Table2

Compound	$EC_{50}(\mu M)^a$	$CC_{50}(\mu M)^b$	$SI^c$	$IC_{50}(\mu M)^d$	
2	NA	$30.1 \pm 2.48$			
3	$3.25 \pm 2.65$	$39.38 \pm 7.25$	12	$28.4 \pm 24.4$	
4	NA.	$23.5 \pm 2.2$			
5	$0.99 \pm 0.35$	$30.9 \pm 10.7$	31	$40.2 \pm 8.6$	
9	$0.391 \pm 0.33$	$276 \pm 52$	340		
15	$0.219 \pm 0.043$	$162 \pm 8.8$	741		
16	$1.15 \pm 0.82$	$79.1 \pm 72.2$	69		
17	$0.068 \pm 0.044$	$30.9 \pm 3.2$	454	$0.83 \pm 0.00$	
20	<b>NA</b>	>275			
21	$0.065 \pm 0.031$	$192 \pm 18.5$	2950		
22	$0.329 \pm 0.175$	$24.5 \pm 10.5$	75		
23	$0.273 \pm 0.174$	$27.6 \pm 8.5$	101		
26	$1.12 \pm 0.22$	> 320	> 286		
27	$0.391 \pm 0.33$	$209 \pm 42$	535		
28	$5.92 \pm 3.69$	$130 \pm 35.8$	22		
29	$0.128 \pm 0.019$	$27.9 \pm 3.2$	220		
32	$0.837 \pm 0.23$	$144.8 \pm 12.1$	173		
33	$0.469 \pm 0.219$	> 313	>667	$9.70 \pm 0.94$	
35	$0.338 \pm 0.246$	$158.5 \pm 51.4$	489		

Anti-HIV-1 activity, cytotoxicity and selectivity index in CEM cells and RT inhibitory activity for some 2,3-diaryl-1,3-thiazolidin-4-ones

<sup>a</sup> 50% Effective concentration or concentration required to protect CEM cells against the cytopathicity of HIV-1–(III<sub>B</sub>) by 50%.<br><sup>b</sup> Cytotoxic concentration or concentration required to reduce CEM cell viability by 50%.

<sup>c</sup> Selectivity index: Ratio CC<sub>50</sub>/EC<sub>50</sub>.<br><sup>d</sup> Poly(C)/oligo(dG) was used as the template/primer and [<sup>3</sup>H]dGTP as the radiolabelled substrate. NA = not active

solution of sodium hydrogen carbonate. After removal of the solvent under reduced pressure, the oily residue was powdered by treatment with a mixture of ethanol and diethyl ether to afford compounds **4–5, 8–9, 11–12, 14–17, 20–23, 26–30, 33** and **35**. The remaining compounds were isolated by silica gel column chromatography eluting with  $CHCl<sub>3</sub>$ (**2–3, 10** and **34**), CHCl3/MeOH 99:1 (**6, 18** and **32**), CHCl3/MeOH 98:2 (**35**) or cyclohexane/EtOAc 70:30 (**24**). All compounds were recrystallized from EtOH. Data of compounds **1, 7, 13, 19, 25** and **31** have been previously reported [\[10–12\].](#page-6-0)

*3.1.1.1. 2-(2,6-Dimethylphenyl)-3-(3-methylphenyl)-1,3-thiazolidin-4-one (2)*. Yield: 35%, m.p.: 103–105 °C; <sup>1</sup> H NMR ( $\delta$ ) 2.26, 2.38 and 2.53 (3s, 9H, Me), 3.95 (s, 2H, CH<sub>2</sub>-5), 6.80–7.15 (m, 8H, ArH and H-2). Anal. ( $C_{18}H_{19}NOS$ ) C, H, N.

*3.1.1.2. 2-(2-Fluoro,6-methoxyphenyl)-3-(3-methylphenyl)- 1,3-thiazolidin-4-one (3)*. Yield: 32%, m.p.: 102–104°C; <sup>1</sup> H NMR (δ) 2.27 (s, 3H, Me), 3.80–4.10 (m, 5H, CH<sub>2</sub>-5 and OMe), 6.60–7.20 (m, 8H, ArH and H-2). Anal.  $(C_{17}H_{16}FNO_2S)$  C, H, N.

*3.1.1.3. 2-(3-Chloro,2-fluoro,6-trifluoromethylphenyl)-3-(3 methylphenyl)-1,3-thiazolidin-4-one (4)*. Yield: 83%, m.p.: 127–129 °C; <sup>1</sup> H NMR (*d*) 2.28 (s, 3H, Me), 3.88 (d, 1H,  $J = 15.4$  Hz, 5-H<sub>A</sub>), 4.13 (dd, 1H,  $J = 1.9$  and 15.4 Hz, 5-H<sub>B</sub>), 6.52 (d, 1H, *J* = 1.9 Hz, H-2), 6.98–7.46 (m, 6H, ArH). Anal.  $(C_{17}H_{12}CIF<sub>4</sub>NOS) C, H, N.$ 

*3.1.1.4. 2-(2-Chloro,6-fluoro,3-methylphenyl)-3-(3-methylphenyl)-1,3-thiazolidin-4-one (5)*. Yield: 85%, m.p.: 130– 133 °C; <sup>1</sup>H NMR ( $\delta$ ) 2.22 and 2.28 (2s, 6H, Me), 3.86 (d, 1H,  $J = 15.4$  Hz,  $5-H_A$ ),  $4.12$  (dd,  $1H, J = 1.9$  and  $15.4$  Hz,  $5-H_B$ ), 6.78 (d, 1H, *J* = 1.9 Hz, H-2), 6.87–7.20 (m, 6H, ArH). Anal.  $(C_{17}H_{15}CIFNOS) C, H, N.$ 

*3.1.1.5. 2-(2,6-Dimethoxyphenyl)-3-(3-methylphenyl)-1,3 thiazolidin-4-one (6)*. Yield: 24%, m.p.: 130–135 °C; <sup>1</sup> H NMR (*d*) 2.25 (s, 3H, Me), 3.73 and 3.91 (2s, 6H, OMe), 3.80 (d, 1H,  $J = 15.1$  Hz,  $5-H_A$ ), 4.05 (dd, 1H,  $J = 1.9$  and 15.1 Hz, 5-H<sub>B</sub>), 6.39–7.16 (m, 8H, ArH and H-2). Anal.  $(C_{18}H_{19}NO_3S)$  C, H, N.

*3.1.1.6. 2-(2,6-Dimethylphenyl)-3-(4-methylpyridin-2-yl)- 1,3-thiazolidin-4-one (8)*. Yield: 33%, m.p.: 110–113 °C; <sup>1</sup> H NMR (*d*) 2.33, 2.41 and 2.59 (3s, 9H, Me), 3.98 (s, 2H, CH2-5), 6.79–8.05 (m, 7H, ArH and H-2). Anal.  $(C_{18}H_{18}N_2OS)$  C, H, N.

*3.1.1.7. 2-(2-Fluoro,6-methoxyphenyl)-3-(4-methylpyridin-2-yl)-1,3-thiazolidin-4-one (9)*. Yield: 28%, m.p.: 136– 139 °C; <sup>1</sup> H NMR (*d*) 2.32 (s, 3H, Me), 3.81 (d, 1H,  $J = 15.7$  Hz, 5-H<sub>A</sub>), 3.88 (s, 3H, OMe), 4.15 (dd, 1H,  $J = 1.4$  and 15.7 Hz, 5-H<sub>B</sub>), 6.56–8.11 (m, 7H, ArH and H-2). Anal.  $(C_{17}H_{15}FN_2O_2S)$  C, H, N.

*3.1.1.8. 2-(3-Chloro,2-fluoro,6-trifluoromethylphenyl)-3-(4 methylpyridin-2-yl)-1,3-thiazolidin-4-one (10)*. Yield: 81%, m.p.: 125–127 °C; <sup>1</sup>H NMR (δ) 2.35 (s, 3H, Me), 3.85 (d, 1H,  $J = 15.9$  Hz,  $5-H_A$ ), 4.21 (dd, 1H,  $J = 1.6$  and 15.9 Hz,  $5-H_B$ ), 6.82–8.01 (m, 6H, ArH and H-2). Anal.  $(C_{17}H_{11}CIF_4N_2OS)$ C, H, N.

*3.1.1.9. 2-(2-Chloro,6-fluoro,3-methylphenyl)-3-(4-methylpyridin-2-yl)-1,3-thiazolidin-4-one (11)*. Yield: 83%, m.p.: 145–148 °C; <sup>1</sup>H NMR ( $\delta$ ) 2.14 and 2.34 (2s, 6H, Me), 3.85 (d, 1H,  $J = 15.7$  Hz,  $5-H_A$ ), 4.19 (dd, 1H,  $J = 1.6$  and 15.7 Hz, 5-H<sub>B</sub>), 6.82–8.10 (m, 6H, ArH and H-2). Anal.  $(C_{17}H_{14}CIFN_2OS) C, H, N.$ 

*3.1.1.10. 2-(2,6-Dimethoxyphenyl)-3-(4-methylpyridin-2 yl)-1,3-thiazolidin-4-one (12)*. Yield: 32%, m.p.: 144– 146 °C; <sup>1</sup> H NMR (*d*) 2.30 (s, 3H, Me), 3.76–3.90 (m, 7H, OMe and  $5-H_A$ ), 4.13 (dd, 1H,  $J = 1.6$  and 15.4 Hz,  $5-H_B$ ), 6.45–8.10 (m, 7H, ArH and H-2). Anal.  $(C_{18}H_{18}N_2O_3S)$  C, H, N.

*3.1.1.11. 2-(2,6-Dimethylphenyl)-3-(6-methylpyridin-2-yl)- 1,3-thiazolidin-4-one (14)*. Yield: 43%, m.p.: 143–146 °C; <sup>1</sup> <sup>1</sup>H NMR ( $\delta$ ) 2.27, 2.39 and 2.63 (3s, 9H, Me), 3.97 (s, 2H, CH2-5), 6.80–7.69 (m, 7H, ArH and H-2). Anal.  $(C_{18}H_{18}N_2OS)$  C, H, N.

*3.1.1.12. 2-(2-Fluoro,6-methoxyphenyl)-3-(6-methylpyridin-2-yl)-1,3-thiazolidin-4-one (15)*. Yield: 34%, m.p.: 113– 115 °C; <sup>1</sup> H NMR (*d*) 2.32 (s, 3H, Me), 3.82 (d, 1H,  $J = 15.7$  Hz, 5-H<sub>A</sub>), 3.86 (s, 3H, OMe), 4.17 (dd, 1H,  $J = 1.6$  and 15.7 Hz, 5-H<sub>B</sub>), 6.58–7.97 (m, 7H, ArH and H-2). Anal. ( $C_{17}H_{15}FN_2O_2S$ ) C, H, N.

*3.1.1.13. 2-(3-Chloro,2-fluoro,6-trifluoromethylphenyl)-3- (6-methylpyridin-2-yl)-1,3-thiazolidin-4-one (16)*. Yield: 86%, m.p.: 125–127 °C; <sup>1</sup> H NMR (*d*) 2.22 (s, 3H, Me), 3.85 (d, 1H,  $J = 15.9$  Hz,  $5-H_A$ ), 4.20 (dd, 1H,  $J = 1.6$  and 15.9 Hz, 5-H<sub>B</sub>), 6.84–8.02 (m, 6H, ArH and H-2). Anal.  $(C_{17}H_{11}ClF_4N_2OS) C, H, N.$ 

*3.1.1.14. 2-(2-Chloro,6-fluoro,3-methylphenyl)-3-(6-methylpyridin-2-yl)-1,3-thiazolidin-4-one (17)*. Yield: 76%, m.p.: 127–128°C; <sup>1</sup> H NMR (*d*) 2.15 and 2.27 (2s, 6H, Me), 3.85 (d, 1H,  $J = 15.7$  Hz,  $5-H_A$ ), 4.21 (dd, 1H,  $J = 1.1$  and 15.7 Hz, 5-H<sub>B</sub>), 6.97–8.29 (m, 6H, ArH and H-2). Anal.  $(C_{17}H_{14}CIFN_2OS) C, H, N.$ 

*3.1.1.15. 2-(2,6-Dimethoxyphenyl)-3-(6-methylpyridin-2 yl)-1,3-thiazolidin-4-one (18)*. Yield: 25%, m.p.: 143– 145 °C; <sup>1</sup>H NMR ( $\delta$ ) 2.30 (s, 3H, Me), 3.78 and 3.83 (2s, 6H, OMe), 3.84 (d, 1H,  $J = 15.4$  Hz,  $5-H_A$ ), 4.15 (dd, 1H,  $J = 1.6$  and 15.4 Hz, 5-H<sub>B</sub>), 6.45–7.86 (m, 7H, ArH and H-2). Anal.  $(C_{18}H_{18}N_2O_3S)$  C, H, N.

*3.1.1.16. 3-(6-Bromopyridin-2-yl)-2-(2,6-dimethylphenyl)- 1,3-thiazolidin-4-one (20)*. Yield: 32%, m.p.: 159–162 °C; <sup>1</sup> <sup>1</sup>H NMR ( $\delta$ ) 2.36 and 2.66 (2s, 6H, Me), 3.98 (s, 2H, CH<sub>2</sub>), 6.82–7.92 (m, 7H, ArH and H-2). Anal.  $(C_{17}H_{15}BrN_2OS)$  C, H, N.

*3.1.1.17. 3-(6-Bromopyridin-2-yl)-2-(2-fluoro,6-methoxyphenyl)-1,3-thiazolidin-4-one (21)*. Yield: 52%, m.p.: 110– 112 °C; <sup>1</sup>H NMR ( $\delta$ ) 3.81 (d, 1H, *J* = 15.9 Hz, 5-H<sub>A</sub>), 3.90 (s, 3H, OMe), 4.18 (dd, 1H,  $J = 1.6$  and 15.9 Hz, 5-H<sub>B</sub>), 6.63– 8.25 (m, 7H, ArH and H-2). Anal.  $(C_{16}H_{12}BrFN_2O_2S)$  C, H, N.

*3.1.1.18. 3-(6-Bromopyridin-2-yl)-2-(3-chloro,2-fluoro,6 trifluoromethylphenyl)-1,3-thiazolidin-4-one (22)*. Yield: 75%, m.p.: 149–151 °C; <sup>1</sup> H NMR (*d*) 3.84 (d, 1H, *J* = 15.9, 5-H<sub>A</sub>), 4.20 (dd, 1H,  $J = 1.6$  and 15.9 Hz, 5-H<sub>B</sub>), 6.98–8.28 (m, 6H, ArH and H-2). Anal.  $(C_{16}H_8BrClF_4N_2OS)$  C, H, N.

*3.1.1.19. 3-(6-Bromopyridin-2-yl)-2-(2-chloro,6-fluoro,3 methylphenyl)-1,3-thiazolidin-4-one (23)*. Yield: 64%, m.p.: 128–129 °C; <sup>1</sup> H NMR (*d*) 2.33 (s, 3H, Me), 3.84 (d, 1H,  $J = 15.9$  Hz,  $5-H_A$ ),  $4.20$  (dd,  $1H, J = 1.6$  and  $15.9$  Hz,  $5-H_B$ ), 6.82–8.05 (m, 6H, ArH and H-2). Anal.  $(C_{16}H_{11}BrCIFN_2OS)$ C, H, N.

*3.1.1.20. 3-(6-Bromopyridin-2-yl)-2-(2,6-dimethoxyphenyl)-1,3-thiazolidin-4-one (24)*. Yield: 23%, m.p.: 126– 129 °C; <sup>1</sup>H NMR ( $\delta$ ) 3.46–3.95 (m, 7H, OMe and 5-H<sub>A</sub>), 4.15 (dd, 1H,  $J = 1.6$  and 15.7 Hz, 5-H<sub>B</sub>), 6.51–8.20 (m, 7H, ArH and H-2). Anal.  $(C_{17}H_{15}BrN_2O_3S)$  C, H, N.

*3.1.1.21. 2-(2,6-Dimethylphenyl)-3-(4,6-dimethylpyridin-2 yl)-1,3-thiazolidin-4-one (26)*. Yield: 42%, m.p.: 155 °C dec.; <sup>1</sup> H NMR (*d*) 2.23, 2.27, 2.40 and 2.62 (4s, 12H, Me), 3.96 (s, 2H, CH<sub>2</sub>), 6.65–7.46 (m, 6H, ArH and H-2). Anal.  $(C_{19}H_{20}N_2OS)$  C, H, N.

*3.1.1.22. 3-(4,6-Dimethylpyridin-2-yl)-2-(2-fluoro,6-methoxyphenyl)- 1,3-thiazolidin-4-one (27)*. Yield: 33%, m.p.: 139–141 °C; <sup>1</sup> H NMR (*d*) 2.26 (s, 6H, Me), 3.81 (d, 1H,  $J = 15.4$  Hz, 5-H<sub>A</sub>), 3.87 (s, 3H, OMe), 4.16 (dd, 1H,  $J = 1.6$  and 15.4 Hz, 5-H<sub>B</sub>), 6.57–7.76 (m, 6H, ArH and H-2). Anal. ( $C_{18}H_{17}FN_2O_2S$ ) C, H, N.

*3.1.1.23. 2-(3-Chloro,2-fluoro,6-trifluoromethylphenyl)-3- (4,6-dimethylpyridin-2-yl)-1,3-thiazolidin-4-one (28)*. Yield: 58%, m.p.: 144–146 °C; <sup>1</sup> H NMR (*d*) 2.18 and 2.30 (2s, 6H, Me), 3.83 (d, 1H,  $J = 15.9$  Hz, 5-H<sub>A</sub>), 4.19 (dd, 1H,  $J = 1.6$  and 15.9 Hz, 5-H<sub>B</sub>), 6.69–7.82 (m, 5H, ArH and H-2). Anal.  $(C_{18}H_{13}CIF_4N_2OS)$  C, H, N.

*3.1.1.24. 2-(2-Chloro,6-fluoro,3-methylphenyl)-3-(4,6-dimethylpyridin-2-yl)-1,3-thiazolidin-4-one (29)*. Yield: 75%, m.p.: 110–113 °C; <sup>1</sup>H NMR (δ) 2.15 and 2.28 (2s, 9H, Me), 3.84 (d, 1H,  $J = 15.9$  Hz,  $5-H_A$ ), 4.21 (dd, 1H,  $J = 1.6$  and 15.9 Hz, 5-H<sub>B</sub>), 6.67–7.86 (m, 5H, ArH and H-2). Anal.  $(C_{18}H_{16}CIFN_2OS) C, H, N.$ 

*3.1.1.25. 2-(2,6-Dimethoxyphenyl)-3-(4,6-dimethylpyridin-2-yl)-1,3-thiazolidin-4-one (30)*. Yield: 28%, m.p.: 158– 160 °C; <sup>1</sup> H NMR (*d*) 2.24 and 2.26 (2s, 6H, Me), 3.77–3.90

<span id="page-5-0"></span>(m, 7H, OMe and 5-H<sub>A</sub>), 4.14 (dd, 1H,  $J = 1.6$  and 15.4 Hz, 5-H<sub>B</sub>), 6.45–7.64 (m, 6H, ArH and H-2). Anal.  $(C_{19}H_{20}N_2O_3S)$  C, H, N.

*3.1.1.26. 2-(2,6-Dimethylphenyl)-3-(4-methylpyrimidin-2 yl)-1,3-thiazolidin-4-one (32)*. Yield: 28%, m.p.: 137– 140 °C; <sup>1</sup> H NMR (*d*) 2.36, 2.51 and 2.59 (3s, 9H, Me), 3.99  $(s, 2H, CH<sub>2</sub>), 6.82-8.48$  (m, 6H, ArH and H-2). Anal.  $(C_{17}H_{17}N_3OS)$  C, H, N.

*3.1.1.27. 2-(2-Fluoro,6-methoxyphenyl)-3-(4-methylpyrimidin-2-yl)-1,3-thiazolidin-4-one (33)*. Yield: 32%, m.p.: 148– 150 °C; <sup>1</sup> H NMR (*d*) 2.38 (s, 3H, Me), 3.84 (d, 1H,  $J = 15.7$  Hz, 5-H<sub>A</sub>), 3.87 (s, 3H, OMe), 4.17 (dd, 1H,  $J = 1.1$  and 15.7 Hz, 5-H<sub>B</sub>), 6.61–8.49 (m, 6H, ArH and H-2). Anal. ( $C_{16}H_{14}FN_3O_2S$ ) C, H, N.

*3.1.1.28. 2-(3-Chloro,2-fluoro,6-trifluoromethylphenyl)-3- (4-methylpyrimidin-2-yl)-1,3-thiazolidin-4-one (34)*. Yield: 23%, m.p.: 148–151 °C; <sup>1</sup> H NMR (*d*) 2.32 (s, 3H, Me), 3.87 (d, 1H,  $J = 15.9$  Hz,  $5-H_A$ ), 4.21 (dd, 1H,  $J = 1.6$  and 15.9 Hz, 5-H<sub>B</sub>), 6.88–8.51 (m, 5H, ArH and H-2). Anal.  $(C_{16}H_{10}CIF_4N_3OS)$  C, H, N.

*3.1.1.29. 2-(2-Chloro,6-fluoro,3-methylphenyl)-3-(4-methylpyrimidin-2-yl)-1,3-thiazolidin-4-one (35)*. Yield: 38%, m.p.: 126 °C dec.; <sup>1</sup>H NMR (δ) 2.17 and 2.39 (2s, 6H, Me), 3.89 (d, 1H,  $J = 15.7$  Hz, 5-H<sub>A</sub>), 4.23 (dd, 1H,  $J = 1.9$  and 15.7 Hz, 5-H<sub>B</sub>), 6.86–8.51 (m, 5H, ArH and H-2). Anal.  $(C_{16}H_{13}CIFN_3OS)$  C, H, N.

*3.1.1.30. 2-(2,6-Dimethoxyphenyl)-3-(4-methylpyrimidin-2 yl)-1,3-thiazolidin-4-one (36)*. Yield: 23%, m.p.: 147– 149 °C; <sup>1</sup> H NMR (*d*) 2.34 (s, 3H, Me), 3.47–3.91 (m, 7H, OMe and 5-H<sub>A</sub>), 4.16 (dd, 1H,  $J = 1.4$  and 15.4 Hz, 5-H<sub>B</sub>), 6.48–8.47 (m, 6H, ArH and H-2). Anal. ( $C_{17}H_{17}N_3O_3S$ ) C, H, N.

## *3.2. Pharmacology*

## *3.2.1. In vitro anti-HIV assay*

The methodology of the anti-HIV assays has been previously described [\[15,16\].](#page-6-0) Briefly, MT-4 or CEM cells were infected with HIV-1 ( $III_B$ ) and HIV-2 (ROD) at 100 CCID<sub>50</sub> (50% cell culture infective dose) per ml of cell suspension. Then, 100 ul of the infected cell suspension was added to microtitre plate containing 100 µl of an appropriate dilution of the test compounds. In MT-4 cells, after 5 d of incubation, the number of viable cells was determined. In CEM cells, after 4 d of incubation, HIV-1-induced syncytium formation was recorded. The 50% effective concentration  $(EC_{50})$  and 50% cytotoxic concentration  $(CC_{50})$  were defined as the compound concentrations required to reduce

cell viability (MT-4) or to inhibit virus-induced cytopathicity (CEM) by 50%, or to reduce by 50% the number of viable cells in mock-infected MT-4 and CEM cell cultures, respectively.

# *3.2.2. HIV-1 RT assay*

The procedure for assessing the reverse transcriptase inhibition has been described previously [\[17\].](#page-6-0) The reaction mixture (50 µl) contained 50 mM Tris–HCl (pH 7.8), 5 mM dithiothreitol, 30 mM glutathione, 50 µM EDTA, 150 mM KCl, 5 mM  $MgCl<sub>2</sub>$ , 1.25 µg of bovine serum albumine, an appropriate concentration of the radiolabelled substrate  $[3]$ 0.1 mM poly(C)·oligo(dG) as the template/primer, 0.06% Triton X-100, 10 µl of inhibitor solution (containing various concentrations of compounds), and 1 µl of RT preparation. The reaction mixtures were incubated at 37 °C for 15 min, at which time 100 µl of calf thymus DNA (150  $\mu$ g/ml), 2 ml of Na<sub>4</sub>P<sub>2</sub>O<sub>7</sub> (0.1 M in 1 M HCl), and 2 ml of trichloroacetic acid (10% v/v) were added. The solutions were kept on ice for 30 min, after which the acid-insoluble material was washed and analysed for radioactivity. For the experiments in which 50% inhibitory concentration  $(IC_{50})$  of the test compounds was determined, fixed concentration of 2.5  $\mu$ M [<sup>3</sup>H]dGTP was used.

## **Acknowledgements**

Financial support for this research by Università di Messina (Fondo Ateneo di Ricerca) and Ministero dell'Istruzione, dell'Università e della Ricerca (COFIN 2002) is gratefully acknowledged.

# **References**

- [1] E. De Clercq, New development in anti-HIV chemotherapy, Biochim. Biophys. Acta 1587 (2002) 258–275.
- [2] R. Esnouf, J. Ren, C. Ross, Y. Jones, D. Stammers, D. Stuart, Mechanism of inhibition of reverse transcriptase by non-nucleoside inhibitors, Nat. Struct. Biol. 2 (1995) 303–308.
- [3] J.A. Tavel, K.D. Miller, H. Masur, Guide to major clinical trials in antiretroviral therapy in human immunodeficiency virus-infected patients: protease inhibitors, non-nucleoside reverse transcriptase inhibitors, and nucleoside reverse transcriptase inhibitors, Clin. Inf. Dis. 28 (1999) 643–676.
- [4] F.J. Palella, K.M. Delaney, A.C. Moorman, M.O. Loveless, J. Fuhrer, G.A. Satten, et al., Declining morbidity and mortality among patients advanced human immunodeficiency virus infection. HIV outpatient study investigators, New. Engl. J. Med. 338 (1998) 853–860.
- [5] A. Chimirri, S. Grasso, C. Molica, A.M. Monforte, P. Monforte, M. Zappalà, et al., Structural features and anti-human immunodeficiency virus (HIV) activity of the isomers of 1-(2′,6′-difluorophenyl)- 1*H*,3*H*-thiazolo[3,4*-a*]benzimidazole, a potent non-nucleoside HIV-1 reverse transcriptase inhibitor, Antiv. Chem. Chemother 8 (1997) 363–370.
- [6] A. Chimirri, S. Grasso, A.M. Monforte, P. Monforte, A. Rao, M. Zappalà, et al., Synthesis, structure and in vitro anti-human immunodeficiency virus activity of novel 3-methyl-1*H,3H-thiazolo[3,4 a]benzimidazoles*, Antiv. Chem. Chemother 9 (1998) 431–438.
- <span id="page-6-0"></span>[7] A. Chimirri, S. Grasso, P. Monforte, A. Rao, M. Zappalà, A.M. Monforte, et al., Synthesis and biological activity of novel 1*H,3Hthiazolo[3,4-a]benzimidazoles: non-nucleoside human immunodeficiency virus type 1 reverse transcriptase inhibitors*, Antiv. Chem. Chemother 10 (1999) 211–217.
- [8] M.L. Barreca, A. Chimirri, L. De Luca, A.M. Monforte, P. Monforte, A. Rao, et al., Discovery of 2,3-diaryl-1,3-thiazolidin-4-ones as potent anti-HIV-1 agents, Bioorg. Med. Chem. Lett. 11 (2001) 1793– 1796.
- [9] A. Rao, A. Carbone, A. Chimirri, E. De Clercq, A.M. Monforte, P. Monforte, et al., Synthesis and anti-HIV activity of 2,3-diaryl-1,3 thiazolidin-4-(thi)one derivatives, Il Farmaco 57 (2002) 747–751.
- [10] M.L. Barreca, J. Balzarini, A. Chimirri, E. De Clercq, L. De Luca, H.D. Höltje, et al., Design, synthesis, structure–activity relationships and molecular modeling studies of 2,3-diaryl-1,3-thiazolidin-4-ones as potent anti-HIV agents, J. Med. Chem. 45 (2002) 5410–5413.
- [11] A. Rao, A. Carbone, A. Chimirri, E. De Clercq, A.M. Monforte, P. Monforte, et al., Synthesis and anti-HIV activity of 2,3-diaryl-1,3 thiazolidin-4-ones, Il Farmaco 58 (2003) 115–120.
- [12] A. Rao, J. Balzarini, A. Carbone, A. Chimirri, E. De Clercq, A.M. Monforte, et al., 2-(2,6-Dihalophenyl)-3-(pyrimidin-2-yl)-1,3 thiazolidin-4-ones as non-nucleoside HIV-1 reverse transcriptase inhibitors, submitted for publication.
- [13] A. Casimiro-Garcia, M. Micklatcher, J.A. Turpin, T.L. Stup, K. Watson, R.W. Buckheit, et al., Novel modifications in the alkenyldiarylmethane (ADAM) series of non-nucleoside reverse transcriptase inhibitors, J. Med. Chem. 42 (1999) 4861–4874.
- [14] J. Ding, K. Das, H. Moereels, L. Koymans, K. Andries, P.A.J. Janssen, et al., Structure of HIV-1 RT/TIBO R 86183 complex reveals similarity in the binding of diverse nonnucleoside inhibitors, Nat. Struct. Biol. 2 (1995) 407–415.
- [15] R. Pauwels, J. Balzarini, M. Baba, R. Snoeck, D. Schols, P. Herdewijin, et al., Rapid and automated tetrazolium-based colorimetric assay for the detection of anti-HIV compounds, J. Virol. Method. 20 (1988) 309–321.
- [16] J. Balzarini, A. Karlsson, M.-J. Pérez-Pérez, L. Vrang, J. Walbers, H. Zhang, et al., HIV-1-specific reverse transcriptase inhibitors show differential activity against HIV-1 mutant strains containing different amino acid substitution in the reverse transcriptase, Virology 192 (1993) 246–253.
- [17] J. Balzarini, M.-J. Pérez-Pérez, A. San-Félix, M.J. Camarasa, I.C. Bathurst, P.J. Barr, et al., Kinetics of inhibition of human immunodeficiency virus type 1 (HIV-1) reverse transcriptase by the novel HIV-1-specific nucleoside analogue [2′,5′-bis-*O*-(*tert*butyldimethylsilyl)-b-D-ribofuranosyl]-3′-spiro-5″-(4″-amino-1″,2″ oxathiole-2″,2″-dioxide)thymine (TSAO-T), J. Biol. Chem. 267 (1992) 11831–11838.